P-5

Unique features and function of ER α-1,2-mannosidase homologues in the human fungal pathogen Cryptococcus

neoformans

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N-glycans play important roles in quality coatrol (QC) of glycoprotein folding in the endoplasmic reticulum (ER) and in ER-associated degradation (ERAD). ER a 1.2-mannosidase I (MNSI) contributes to ERQC and ERAD by trimming a mannose residue from the B branch of the N-oligosaccharides, initiating the formation of the molecular signals that either lead folded proteins towards the Golgi apparatus, or direct misfolded glycoproteins towards degradation in the cytoplasm (1). The human-pathogen Cryptococcus neoformans has a unique N-glycosylation lacking ER-associated glucosyltransferases but carrying multiple mannosidases (2). To investigate the molecular assembly of C. neoformans specific-glycoprotein QC pathway and its impact on pathogenicity, we disrupted two genes encoding putative ER \(\alpha\)-1,2-mannosidases, MNSIA and MNSIB. The N-glycan profiles suggested that, whereas Mns1Ap acts as the widely known ER-α-mannosidase I, Mns1Bp appears to be a novel mannosidase involved in mannose processing in the Golgi. Interestingly, both mannosidases were shown to localize mostly in the Golgi apparatus, as evidenced by the presence of punctate fluorescence patterns resembling golgi stacks. Although the mns1AD mutant strain showed only mild growth retardation under several stress conditions, the mns1BD displayed the increased sensitivity to high temperature at 39°C and notably increased resistance to SDS. The double mutant strain (mns1ADmns1BD) exhibited slightly increased sensitivity to fludioxonil treatment. Moreover, a considerable reduction of capsule mass was observed in both mns1AD and mns1BD, with the most severe defect in the mns1ADmns1BD mutant, suggestive of association with in vivo virulence. Overall, our data strongly suggest that the functions of both MNS1A and MNS1B are involved in proper N-glycan processing for glycoprotein QC, which is required for capsule assembly and attachment in C. neoformans.

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